

Specifications

■ Microscope

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| Microscope configuration | transmission inverted microscope |
| Microscopy techniques | holography (quantitative phase imaging), epifluorescence, simulated DIC, brightfield, high-pass filtered phase |
| Objectives | magnification 4× to 60× |
| Objective turret | 6-position, motorized exchange |
| Light source | halogen lamp |
| Operating wavelength | 650 nm |
| Sample stage | motorized, 130 mm × 90 mm travel range |
| Focusing | motorized objective turret, 8 mm travel range |
| Piezo-focusing | optional, travel range 500 μm |
| Lateral resolution | 3.3 μm with 4× NA 0.1 objective 0.57 μm with 60× NA 1.4 objective |
| Field of view | objective dependent, up to 1.7 mm × 1.7 mm with 4× objective |
| Acquisition framerate | 5.5 fps at full frame (option: higher framerates possible) |
| Reconstructed phase image size | 1200 px × 1200 px |
| Illumination power at sample plane | down to 0.2 μW/cm ² |
| Phase detection sensitivity | down to 0.0035 rad (0.7 nm at Δn = 0.5) Δn - difference between refractive indexes of sample and surrounding media |
| Power | 230 V/50 Hz (120 V/60 Hz optional), 1200 VA |
| Dimensions (W × L × H) | 1100 mm × 950 mm × 1620 mm microscope with incubator 2515 mm × 974 mm × 1620 mm total with operator table |
| Weight | 350 kg (including microscope table, fluorescence module and microscope incubator) |

Field and aperture diaphragms

Side port available for fluorescence module or other additional techniques

Microscope table with anti-vibration suspension

Control panel with multifunctional touchscreen, sample stage joystick and rotary knobs

Microscope incubator with computer temperature setting and temperature data logging (optional)

Incubation chamber for precise and long-term control of temperature, humidity and CO₂ concentrations (optional)

■ Fluorescence module (optional)

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| Light engines | Lumencor with 3 channels (optionally up to 5 channels) |
| Detectors | standard CCD 1.4 Mpix (1392 px × 1040 px) optional high-sensitivity sCMOS 5.5 Mpix (2560 px × 2160 px) |
| Filters | 3 multichannel filter cubes, motorized channel switching |

Q-PHASE users

■ University of North Florida & Mayo Clinic, Jacksonville, USA

- Cancer research

■ Max Planck Institute of Molecular Cell Biology and Genetics (MPI-CBG), Dresden, Germany

- Quantitative analysis of protein droplets, mouse skull progenitors, growth & degrowth in planarian flatworms

■ Masaryk University Brno, Czech Republic, Faculty of Medicine, Department of Pathological Physiology

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■ Brno University of Technology, Experimental Biophotonics Group

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- J. Collakova, et al.: Coherence-controlled holographic microscopy enabled recognition of necrosis as the mechanism of cancer cells death after exposure to cytopathic turbid emulsion, *J. Biomed. Opt.* 20(11), 2015.
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- A. Krizova, et al.: Dynamic phase differences based on quantitative phase imaging for the objective evaluation of cell behavior, *J. Biomed. Opt.* 20(11), 2015.
- M. Lostak, et al.: Coherence-controlled holographic microscopy in diffuse media, *Opt. Express* 22(4), 2014.
- H. Janeckova, et al.: Proving Tumour Cells by Acute Nutritional/Energy Deprivation as a Survival Threat: A Task for Microscopy, *Anticancer Res.* 29(6), 2009.

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- Osmotic changes in cells, cell reaction to treatment, cells in 3D environment
- L. Pastorek, et al.: Holography microscopy as an artifact-free alternative to phase-contrast, *Histochem Cell Biol.* 149(2), 2018.

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